

Chapter 12

Anti-Infective Agents and Antimicrobial Stewardship

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Abstract: Infectious diseases remain a formidable threat to global health, despite substantial advances in modern medicine. The extensive use and misuse of antimicrobial agents have led to the emergence of resistant pathogens, posing significant challenges in treatment. This chapter provides an in-depth analysis of anti-infective pharmacology, encompassing antibacterial, antiviral, antifungal, and antiparasitic agents. It outlines their mechanisms of action, spectrum of activity, and clinical applications. Special emphasis is placed on the molecular underpinnings of antimicrobial resistance, including enzymatic degradation, efflux pumps, and biofilm formation. The chapter also addresses strategies to mitigate resistance, highlighting antimicrobial stewardship programs (ASPs), diagnostic innovations, and the development of novel therapeutics such as phage therapy and new beta-lactamase inhibitors. By integrating evidence-based approaches with practical implementation, this chapter underscores the critical need for precision in prescribing practices and institutional policies. Furthermore, it reviews global regulatory efforts like the WHO AWaRe classification system to guide rational antimicrobial use. Through a synthesis of current research, comparative insights, and forward-looking perspectives, this chapter equips healthcare professionals with the foundational and emerging tools necessary to combat infectious diseases in the era of resistance.

Keywords: antimicrobial resistance, antibacterial agents, antiviral therapy, antimicrobial stewardship, novel anti-infectives

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12.0 INTRODUCTION

The 21st century has witnessed tremendous progress in antimicrobial discovery and infectious disease control; however, the global burden of infections remains unrelenting. Infectious diseases account for approximately 25% of all deaths worldwide, with an even higher impact in low- and middle-income countries where access to effective therapy is limited [1]. The advent of antimicrobial agents revolutionized medicine, transforming once-lethal infections into manageable conditions. Yet, their indiscriminate and prolonged use has fostered the development of antimicrobial resistance (AMR), now recognized by the World Health Organization (WHO) as one of the top ten global public health threats [2].

The pharmacologic arsenal against infectious agents includes antibiotics, antivirals, antifungals, and antiparasitics each targeting specific classes of pathogens with unique biochemical pathways. These agents operate through distinct mechanisms, such as inhibiting cell wall synthesis, disrupting nucleic acid replication, or impeding protein synthesis. Despite their success, the emergence of multidrug-resistant organisms (MDROs) such as carbapenem-resistant Enterobacteriaceae, methicillin-resistant *Staphylococcus aureus* (MRSA), and extended-spectrum β -lactamase (ESBL) producers have curtailed the efficacy of many standard therapies [3].

This chapter explores anti-infective agents comprehensively, starting with a mechanistic understanding of their pharmacology and moving toward current trends in clinical use, resistance patterns, and stewardship strategies. It delves into both traditional antimicrobial classes and novel therapeutic approaches that are reshaping the future of infectious disease treatment. Special attention is paid to antimicrobial stewardship, diagnostic biomarkers, and regulatory initiatives that promote evidence-based use and development of new antimicrobials.

In the face of growing microbial resistance and limited drug development pipelines, optimizing the use of existing anti-infective agents while advancing new treatment paradigms is paramount. The integration of pharmacological knowledge, microbial surveillance, and institutional stewardship will be the cornerstone of future infectious disease control.

12.1 Antibacterial Agents

The foundation of antimicrobial therapy lies in the use of antibacterial agents, which have been central to infection management since the discovery of penicillin in 1928. These agents target bacterial-specific structures and metabolic processes, including the cell wall, protein synthesis machinery, nucleic acid replication, and folate metabolism. Depending on their spectrum of activity and bacteriostatic or bactericidal nature, antibiotics are used in various clinical scenarios, ranging from empiric therapy to targeted treatment based on culture sensitivity.

12.1.1 Cell Wall, Protein Synthesis, and DNA Inhibitors

Beta-lactams such as penicillins and cephalosporins function by inhibiting transpeptidases penicillin-binding proteins essential for bacterial cell wall synthesis. These agents are highly effective against Gram-positive organisms and, with certain modifications, Gram-negative species as well [4]. Cephalosporins are further subclassified into generations with varying spectra; for instance, ceftriaxone is widely used for community-acquired pneumonia, while cefepime exhibits strong activity against *Pseudomonas aeruginosa*.

Protein synthesis inhibitors such as macrolides (e.g., azithromycin), aminoglycosides (e.g., gentamicin), and tetracyclines (e.g., doxycycline) act on the bacterial ribosome selectively targeting 30S or 50S subunits, thereby impairing mRNA translation. These agents are indispensable in treating

respiratory tract infections, sexually transmitted diseases, and certain atypical pathogens like *Mycoplasma pneumoniae* and *Chlamydia trachomatis* [5].

Fluoroquinolones, such as ciprofloxacin and levofloxacin, target bacterial DNA gyrase and topoisomerase IV, disrupting replication and transcription. Their broad-spectrum activity, excellent oral bioavailability, and tissue penetration make them valuable in treating urinary tract infections, gastrointestinal infections, and certain nosocomial infections. However, increasing resistance has limited their empiric use [6].

Aminoglycosides remain important for severe Gram-negative infections and are often used in synergy with beta-lactams for enhanced efficacy. However, their nephrotoxicity and ototoxicity necessitate careful monitoring [7].

The choice of antibacterial therapy must consider pathogen susceptibility, infection site, pharmacokinetic-pharmacodynamic (PK-PD) parameters, and patient-specific factors. Advances in microbiological diagnostics have enabled tailored therapy, though empirical treatment is often necessary in critical care settings, pending culture results.

Table 12.1: Mechanisms of Action and Examples of Major Antibacterial Classes

Antibacterial Class	Mechanism of Action	Target Site	Representative Agents
Beta-lactams	Inhibition of cell wall synthesis	Penicillin-binding proteins (PBPs)	Penicillin, Cephalosporins, Carbapenems
Aminoglycosides	Inhibition of protein synthesis (30S subunit)	Ribosome	Gentamicin, Amikacin
Macrolides	Inhibition of protein synthesis (50S subunit)	Ribosome	Azithromycin, Erythromycin
Fluoroquinolones	Inhibition of DNA replication	DNA gyrase, Topoisomerase IV	Ciprofloxacin, Levofloxacin
Glycopeptides	Inhibition of peptidoglycan polymerization	Cell wall	Vancomycin
Tetracyclines	Inhibition of protein synthesis (30S subunit)	Ribosome	Doxycycline, Minocycline
Sulfonamides + Trimethoprim	Inhibition of folic acid synthesis	Dihydropteroate & dihydrofolate reductase	Co-trimoxazole

12.2 Antivirals

Viruses present unique therapeutic challenges due to their reliance on host cellular machinery for replication. Consequently, antiviral drugs are designed to target specific steps in the viral life cycle while minimizing host cell toxicity. Modern antiviral therapy has expanded significantly, especially in response to global viral epidemics such as HIV/AIDS, hepatitis C, and COVID-19.

12.2.1 RNA and DNA Virus Therapies

Antivirals targeting DNA viruses, such as herpes simplex virus (HSV) and varicella-zoster virus (VZV), include nucleoside analogs like acyclovir and valacyclovir. These agents undergo phosphorylation within infected cells to inhibit viral DNA polymerase, preventing genome replication

[8]. Acyclovir is a first-line agent for HSV encephalitis and genital herpes, while its prodrug valacyclovir offers improved bioavailability for outpatient use.

In the context of RNA viruses, remdesivir has gained prominence as a nucleotide analog inhibiting RNA-dependent RNA polymerase. Initially developed for Ebola, remdesivir demonstrated activity against SARS-CoV-2 and received emergency use authorization for COVID-19 management [9]. Other agents such as favipiravir and molnupiravir are also under evaluation for influenza and coronavirus infections.

Antiretroviral therapy (ART) for HIV infection comprises multiple drug classes, including reverse transcriptase inhibitors (e.g., tenofovir, lamivudine), protease inhibitors (e.g., lopinavir), and integrase strand transfer inhibitors (e.g., dolutegravir). Combination ART has transformed HIV from a fatal disease to a manageable chronic condition and significantly reduced viral transmission [10].

For hepatitis B and C viruses (HBV and HCV), direct-acting antivirals (DAAs) have revolutionized treatment. Sofosbuvir, a nucleotide analog, combined with NS5A inhibitors like ledipasvir or velpatasvir, has achieved sustained virologic response rates exceeding 95% in HCV infections [11]. HBV management relies on nucleos(t)ide analogs such as entecavir and tenofovir, which suppress viral replication but do not cure the infection.

Despite these successes, antiviral resistance, drug interactions, and the need for long-term adherence remain significant barriers. Personalized medicine approaches, including resistance testing and therapeutic drug monitoring, are increasingly used to optimize antiviral regimens.

12.3 Antifungals and Antiparasitics

Fungal and parasitic infections, while less common than bacterial or viral infections, pose significant morbidity and mortality risks especially in immunocompromised individuals. Advances in medical technology, such as organ transplantation and intensive chemotherapy, have increased the population susceptible to opportunistic infections, emphasizing the need for potent antifungal and antiparasitic agents.

12.3.1 Echinocandins, Azoles, Antimalarials

Echinocandins, such as caspofungin, micafungin, and anidulafungin, target the synthesis of β -(1,3)-D-glucan, an essential component of the fungal cell wall. These agents exhibit fungicidal activity against *Candida* species and fungistatic effects against *Aspergillus*, making them first-line agents in invasive candidiasis, especially in critically ill patients [12]. Echinocandins are notable for their low toxicity and minimal drug-drug interactions, although their intravenous-only formulation limits outpatient use.

Azoles, including fluconazole, voriconazole, and posaconazole, act by inhibiting the fungal cytochrome P450 enzyme lanosterol 14 α -demethylase, thereby disrupting ergosterol synthesis and membrane integrity. Fluconazole remains effective for mucosal and systemic *Candida albicans* infections, whereas voriconazole is the treatment of choice for invasive aspergillosis [13]. However, emerging resistance among non-*albicans* species and pharmacokinetic variability of voriconazole necessitate therapeutic drug monitoring.

Amphotericin B, a polyene antifungal, binds directly to ergosterol, forming membrane pores that lead to cell death. While effective against a broad range of fungi, its use is limited by nephrotoxicity and infusion-related reactions. Liposomal formulations offer improved safety profiles [14].

Antiparasitic therapies remain crucial in endemic regions, addressing diseases such as malaria, leishmaniasis, and helminthiasis. Antimalarial agents like artemisinin-based combination therapies (ACTs) have become the mainstay of *Plasmodium falciparum* treatment due to their rapid efficacy and resistance suppression [15]. Other antiparasitics, such as albendazole for helminths and miltefosine for visceral leishmaniasis, target unique parasitic structures or metabolic processes, offering specificity and reduced host toxicity.

Resistance among fungal pathogens (e.g., *Candida auris*) and parasites (e.g., chloroquine-resistant *Plasmodium*) is on the rise, demanding vigilant surveillance and ongoing drug development. Innovations in antifungal diagnostics and pharmacogenomics hold promise for tailored therapy in the future.

12.4 Antimicrobial Resistance Mechanisms

Antimicrobial resistance (AMR) arises from genetic mutations, horizontal gene transfer, and selective pressure exerted by antimicrobial exposure. Understanding the molecular mechanisms by which pathogens evade therapy is essential for developing new agents and refining existing treatment strategies.

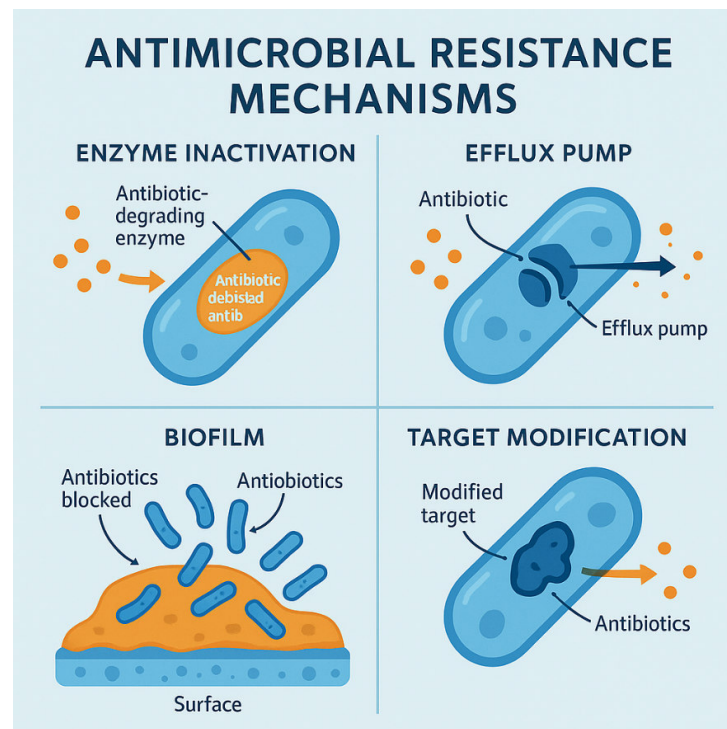


Figure 12.1: Major Mechanisms of Antimicrobial Resistance

12.4.1 Enzymes, Efflux Pumps, Biofilms

A primary mechanism of bacterial resistance is enzymatic degradation of antibiotics. β -lactamases hydrolyze the β -lactam ring in penicillins and cephalosporins, rendering them ineffective. Extended-spectrum β -lactamases (ESBLs) and carbapenemases, such as New Delhi metallo- β -lactamase (NDM), confer resistance to nearly all β -lactam agents, complicating treatment options [16].

The use of β -lactamase inhibitors (e.g., clavulanic acid, avibactam) can restore antibiotic activity in some cases, though resistance to inhibitors is also emerging.

Efflux pumps are another common resistance mechanism. These membrane proteins actively expel antibiotics from the bacterial cell, reducing intracellular drug concentration below therapeutic levels. Efflux-mediated resistance is particularly relevant in tetracyclines, fluoroquinolones, and macrolides [17].

Biofilms structured microbial communities encased in a protective extracellular matrix present a formidable barrier to antimicrobial penetration. Biofilm-associated infections, such as catheter-related bloodstream infections and prosthetic joint infections, are notoriously difficult to treat and often require device removal [18]. Within biofilms, bacteria exhibit phenotypic resistance and reduced metabolic activity, further diminishing drug efficacy.

Other resistance mechanisms include target site modification (e.g., altered penicillin-binding proteins in MRSA), reduced membrane permeability (e.g., porin loss in Gram-negative bacteria), and adaptive resistance driven by stress responses. Genetic elements such as plasmids, transposons, and integrons facilitate the spread of resistance genes among bacterial populations.

Fungal and viral pathogens also develop resistance through distinct mechanisms. In *Candida* species, resistance to azoles is mediated by upregulated efflux transporters and mutations in ERG11, the gene encoding lanosterol demethylase [19]. In viruses, resistance arises from mutations in polymerase or protease genes, as seen in HIV and hepatitis B.

Addressing AMR requires comprehensive strategies involving surveillance, judicious antimicrobial use, infection control, and ongoing research into novel therapeutics and diagnostics.

12.5 Antimicrobial Stewardship Programs (ASPs)

Antimicrobial stewardship programs are structured interventions aimed at optimizing the use of antimicrobial agents to improve clinical outcomes, minimize adverse effects, reduce resistance development, and lower healthcare costs. These programs are now a core component of quality care in hospitals and healthcare systems worldwide.

12.5.1 Implementation and Metrics

ASPs typically involve a multidisciplinary team comprising infectious disease physicians, clinical pharmacists, microbiologists, and infection control specialists. Core strategies include formulary restriction, prospective audit with feedback, dose optimization, de-escalation of broad-spectrum therapy, and adherence to clinical guidelines [20].

One key component is the use of empiric therapy protocols based on local antibiograms, followed by de-escalation to narrower-spectrum agents once culture results become available. This targeted approach reduces unnecessary exposure to broad-spectrum drugs, thereby limiting resistance selection pressure.

Metrics for evaluating ASP performance include days of therapy (DOT) per 1000 patient-days, antimicrobial utilization rates, resistance trends, clinical outcomes (e.g., infection cure rates, mortality), and cost savings. The implementation of electronic decision support tools has improved real-time stewardship by integrating microbiological data, dosing calculators, and guideline alerts into the prescribing process [21].

Educational initiatives form the foundation of sustainable stewardship. Clinicians are trained to interpret microbiology results, understand PK-PD principles, and appreciate the consequences of

inappropriate prescribing. Institutional culture change is also critical, fostering accountability and reinforcing best practices.

Challenges in ASP implementation include resource limitations, lack of trained personnel, and resistance from prescribers. However, studies consistently demonstrate that ASPs reduce antimicrobial use by 20–40% without compromising patient outcomes [22].

In addition to hospital-based programs, antimicrobial stewardship is increasingly being extended to outpatient settings, long-term care facilities, and veterinary medicine. National and global initiatives, such as the CDC's Core Elements of Hospital ASPs and the WHO's Global Action Plan on AMR, provide frameworks for implementation and monitoring.

As AMR continues to threaten modern medicine, ASPs will play an essential role in preserving the effectiveness of existing antimicrobial agents while supporting the integration of novel therapeutics into clinical practice.

12.6 Spectrum of Antimicrobial Use

The concept of antimicrobial spectrum referring to the range of microorganisms against which a drug is effective is central to rational therapy. Antimicrobials are broadly categorized as narrow-spectrum (targeting specific groups) or broad-spectrum (acting against diverse organisms), and their use must be guided by clinical context and microbiological data to maximize efficacy and minimize resistance development.

12.6.1 Narrow vs Broad-Spectrum Therapy

Narrow-spectrum antibiotics, such as penicillin G or clindamycin, are active against a limited subset of organisms and are preferred when the causative pathogen is known. These agents offer the advantage of preserving normal microbiota and reducing selective pressure for resistance among unrelated pathogens. In contrast, broad-spectrum agents such as carbapenems, third-generation cephalosporins, or fluoroquinolones are effective against a wide array of Gram-positive and Gram-negative organisms and are commonly used for empiric therapy in critically ill patients [23].

Empiric therapy is initiated when immediate treatment is necessary, such as in sepsis or meningitis, and culture results are pending. In such cases, broad-spectrum agents are administered to ensure coverage of likely pathogens. However, this approach must be followed by de-escalation based on culture and sensitivity findings. De-escalation to a narrow-spectrum agent reduces collateral damage, including disruption of the microbiome and emergence of multidrug-resistant organisms (MDROs) [24].

The principle of “start smart then focus,” as promoted by the UK's National Health Service (NHS), embodies this approach: begin with appropriate empiric therapy, review at 48–72 hours, and modify based on microbiological data and clinical response. Pharmacokinetic and pharmacodynamic considerations further refine dosing regimens, particularly in critically ill patients with altered drug distribution and clearance.

The inappropriate use of broad-spectrum antibiotics is a major contributor to antimicrobial resistance, with fluoroquinolones and third-generation cephalosporins often implicated. Diagnostic uncertainty, fear of treatment failure, and time pressures are common drivers of this practice.

An evidence-based approach, guided by local resistance patterns (antibiograms), infection severity, patient comorbidities, and clinical judgment, is essential for balancing effective therapy with antimicrobial conservation. Precision antimicrobial use is increasingly supported by stewardship

interventions and diagnostic technologies that enable earlier and more accurate pathogen identification.

Table 12.2: WHO AWaRe Classification of Antibiotics (Selected Examples)

AWaRe Category	Definition	Examples	Recommended Use
Access	First- or second-line agents with lower resistance potential	Amoxicillin, Nitrofurantoin, Doxycycline	Widely available; used for common infections
Watch	Higher resistance potential; critical in stewardship efforts	Ciprofloxacin, Ceftriaxone, Azithromycin	Restricted use; avoid empiric use unless justified
Reserve	Last-resort agents for multidrug-resistant organisms	Colistin, Linezolid, Ceftazidime-avibactam	Only in confirmed resistant infections

12.7 Diagnostics and Biomarkers

Accurate and timely diagnosis is a cornerstone of effective antimicrobial therapy. Diagnostic delays or inaccuracies contribute to inappropriate prescribing, which in turn drives resistance, increases costs, and worsens outcomes. Emerging technologies in rapid diagnostics and biomarkers are reshaping the landscape of infectious disease management.

12.7.1 Rapid Pathogen Identification

Conventional culture methods, while reliable, often require 48–72 hours to yield results. In contrast, rapid diagnostic tools such as matrix-assisted laser desorption/ionization–time of flight (MALDI-TOF) mass spectrometry, polymerase chain reaction (PCR)-based assays, and next-generation sequencing (NGS) offer near-real-time identification of pathogens directly from blood, respiratory, or sterile site specimens [25].

For instance, syndromic panels using multiplex PCR can simultaneously detect bacteria, viruses, and resistance genes from a single sample, often within hours. Such tools improve the accuracy of empiric therapy and facilitate timely de-escalation or escalation of treatment based on definitive results. NGS allows for comprehensive pathogen profiling and outbreak investigation but remains limited by cost and data interpretation complexity.

Biomarkers such as C-reactive protein (CRP) and procalcitonin (PCT) assist in differentiating bacterial from viral infections and gauging treatment response. Procalcitonin levels, in particular, rise in bacterial infections and decline with effective therapy, making it a useful tool for guiding antibiotic initiation and duration in respiratory and sepsis syndromes [26].

In meta-analyses, PCT-guided therapy has been shown to reduce antibiotic exposure without increasing mortality or treatment failure, especially in intensive care and emergency settings. However, biomarker interpretation should always be integrated with clinical assessment, imaging, and microbiological data.

Emerging diagnostics include point-of-care (POC) molecular tests and biosensors that allow decentralized testing in outpatient or low-resource settings. Integration of artificial intelligence and machine learning into diagnostic platforms is also being explored to enhance interpretability and clinical decision-making.

The widespread adoption of rapid diagnostics has the potential to transform antimicrobial use by enabling precision therapy from the outset. Nonetheless, issues such as cost, access, clinician education, and data management need to be addressed to ensure effective implementation.

12.8 Novel Antimicrobials

With the rapid evolution of antimicrobial resistance and a dwindling pipeline of new drugs, the development of novel antimicrobials and innovative therapeutic strategies has become a global priority. Beyond traditional antibiotics, emerging therapies include bacteriophages, novel β -lactamase inhibitors, antimicrobial peptides, and microbiome-based treatments.

12.8.1 Phage Therapy, New Beta-Lactamase Inhibitors

Phage therapy involves the use of bacteriophages viruses that specifically infect and lyse bacteria as targeted antimicrobial agents. Phages offer high specificity, self-amplifying capacity at infection sites, and efficacy against multidrug-resistant bacteria. Clinical case reports and small trials have demonstrated the successful use of phage therapy in treating chronic *Pseudomonas* infections, prosthetic joint infections, and even sepsis caused by *Acinetobacter baumannii* [27].

However, challenges in phage therapy include regulatory hurdles, immune neutralization, limited host range, and the need for individualized preparations. Advances in synthetic biology and phage engineering are addressing these limitations, paving the way for broader clinical application.

New β -lactam/ β -lactamase inhibitor combinations such as ceftazidime-avibactam, meropenem-vaborbactam, and imipenem-relebactam have expanded the therapeutic options for carbapenem-resistant Enterobacteriaceae (CRE) and other ESBL-producing pathogens [28]. These agents inhibit a wider range of β -lactamases, including class A, C, and some class D enzymes, offering effective coverage in previously untreatable infections.

Other novel agents under development include antimicrobial peptides (e.g., daptomycin analogs), monoclonal antibodies targeting bacterial toxins, and agents disrupting bacterial quorum sensing or biofilm formation. Additionally, microbiome restoration therapies using live biotherapeutics (e.g., fecal microbiota transplantation or defined consortia) aim to prevent colonization by resistant pathogens.

Despite these innovations, the economics of antimicrobial development remain unfavorable due to short treatment durations, rapid resistance emergence, and the need for stewardship. Incentives such as the U.S. GAIN (Generating Antibiotic Incentives Now) Act and global funding initiatives are critical to reviving antimicrobial research and development.

12.9 Regulatory Guidelines

Global agencies and health systems have recognized the urgent need for coordinated policies to manage antimicrobial use and resistance. Regulatory frameworks aim to standardize stewardship practices, promote surveillance, and incentivize innovation.

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12.9.1 WHO AWaRe Classification

The World Health Organization's AWaRe classification system categorizes antibiotics into three groups Access, Watch, and Reserve to guide national formularies, monitor consumption, and reduce misuse [29].

- **Access** antibiotics are first- or second-line agents with narrow spectra and lower resistance potential (e.g., amoxicillin, nitrofurantoin). They should be widely available and affordable.
- **Watch** antibiotics (e.g., fluoroquinolones, third-generation cephalosporins) have higher resistance potential and are recommended for specific indications.
- **Reserve** antibiotics, such as colistin or linezolid, are last-resort agents for multidrug-resistant infections and should be used under strict stewardship oversight.

The AWaRe system, endorsed by national health authorities and international organizations, forms the basis for essential medicines lists and procurement policies. It also provides a surveillance framework to track antibiotic usage and resistance trends globally.

Other regulatory initiatives include the U.S. FDA's streamlined approval pathways for priority antimicrobials, the European Medicines Agency's adaptive licensing models, and national action plans aligned with the Global Action Plan on AMR. Surveillance programs such as the WHO's Global Antimicrobial Resistance and Use Surveillance System (GLASS) and the U.S. CDC's National Healthcare Safety Network (NHSN) facilitate the collection of standardized data to inform policy and stewardship.

Regulations also extend to veterinary and agricultural antibiotic use, given the significant contribution of non-human use to resistance. Bans on growth-promoting antibiotics in livestock and the enforcement of prescription-only use are steps toward a One Health approach.

In summary, regulatory frameworks such as AWaRe not only support rational antimicrobial use but also align clinical practice with public health goals, ensuring a sustainable response to AMR.

12.10 CONCLUSION

The global crisis of infectious diseases, compounded by the rapid rise of antimicrobial resistance, represents one of the most pressing challenges in modern medicine. While the discovery of anti-infective agents has revolutionized clinical practice, the persistent misuse and overuse of these therapies have jeopardized their long-term efficacy. This chapter has provided a comprehensive overview of the various classes of anti-infective agents antibacterials, antivirals, antifungals, and antiparasitics highlighting their mechanisms of action, clinical applications, and evolving patterns of

resistance. The molecular basis of antimicrobial resistance ranging from enzymatic degradation and efflux mechanisms to biofilm formation and genetic transmission underscores the complexity of microbial adaptation. Such resistance not only compromises individual patient outcomes but also threatens public health infrastructure, especially in resource-limited settings. Antimicrobial stewardship programs emerge as a vital strategy to preserve existing therapies while ensuring optimal patient care. By promoting evidence-based prescribing, timely de-escalation, and education, these programs serve to balance efficacy with responsibility. The integration of rapid diagnostics, biomarkers such as procalcitonin, and institutional protocols reinforces clinical decision-making and enhances outcomes. Innovative therapeutics, including bacteriophage therapy, next-generation β -lactamase inhibitors, and microbiome-based treatments, offer promising avenues to overcome current therapeutic limitations. However, their success depends on supportive regulatory frameworks and global collaboration. The WHO AWaRe classification exemplifies how international guidelines can influence antibiotic usage patterns, drive stewardship, and shape policy. As the field evolves, the emphasis must shift from reactive to proactive strategies fostering rational drug development, global surveillance, and integrated One Health approaches. The future of anti-infective pharmacology lies in harnessing multidisciplinary tools, fostering global cooperation, and embedding stewardship at the core of healthcare delivery.

In this era of emerging pathogens and waning antibiotic efficacy, clinical prudence, scientific innovation, and coordinated action are not only desirable but essential. The sustainable management of infectious diseases depends not merely on new drugs, but on a transformed understanding of how, when, and why we use them.

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